640

Erythrinaline Alkaloids and Antimicrobial Flavonoids from *Erythrina latissima*

Cornelius C. W. Wanjala^{1,3}, Benard F. Juma¹, Gomotsang Bojase¹, Berhanu A. Gashe², Runner R.T. Majinda¹

Abstract

The seed pods of *Erythrina latissima* yielded erysotrine, erysodine, syringaresinol, vanillic acid, a new erythrina alkaloid, (+)-10,11-dioxoerysotrine, which was lethal to brine shrimp and 2-(5'-hydroxy-3'-methoxy phenyl)-6-hydroxy-5-methoxybenzofuran, which showed strong antimicrobial activity against the yeast spores, Gram-positive and Gram-negative bacteria. The root bark gave four known pterocarpans which showed moderate to strong antifungal activity against the yeast spores and three known flavonoids showed antimicrobial activity against all test microorganisms.

The genus *Erythrina* comprises over 110 species showing cosmopolitan distribution [1] and is known to produce *C*-prenylated flavanones, isoflavanones and pterocarpans [2], [3]. *Erythrina latissima* E. Meyer (Fabaceae) is a tree 9–24 m tall, found in Botswana, Zimbabwe and South Africa and whose root and stem are burnt and used for dressing open wounds [4] while the seeds are known to contain the erythraline alkaloids [5]. No prior phytochemical work has, to date, been reported on the seed pods of this genus.

The HR-TOF-EIMS of the isolated, new erythrina alkaloid 1 showed a molecular ion peak at 341.1265 consistent with the molecular formula $C_{19}H_{19}NO_2$. The 1H - and ^{13}C -NMR data (Table 1) for 1 are very similar to those of (+)-10,11-dioxoerythraline which was reported from E. bidwillii [6] with the only difference that the latter contains a methylenedioxy functional group $(\delta_{\rm H}$ = 5.97, 2H) in place of the two methoxy groups ($\delta_{\rm H}$ = 3.95 and 3.91) in compound 1. The nature and identity of the tetracyclic ring moiety was deduced from the ¹H-NMR data which along with published data [5], [6], [7] enabled us to identify the parent moiety of **1** as (+)-erysotrine. The ¹³C-NMR data for **1** revealed the presence of two carbonyls at $\delta_{\rm C}$ = 159.5 (amide) and $\delta_{\rm C}$ = 181.8 (α , β -unsaturated) and the placement of the former at C-10 and the latter at C-11 was based on HMBC. The rest of the structure was assigned and confirmed using HMBC and other 2D NMR data. Compound 1 is thus a 10,11-dioxo-analogue of ery-

Affiliation: ¹ Department of Chemistry, University of Botswana, Gaborone, Botswana \cdot ² Department of Biological Sciences, University of Botswana, Gaborone, Botswana \cdot ³ Jomo Kenyatta University of Agriculture and Technology, Nairobi, Kenya

Correspondence: Dr. R. R.T. Majinda · Department of Chemistry · University of Botswana · Private Bag 00704, Gaborone, Botswana · E-Mail: majindar@mopipi.ub.bw · Fax: +267-355 2836

Received: October 2, 2001 · Accepted: January 12, 2002

Bibliography: Planta Med 2002; 68: 640–642 · © Georg Thieme Verlag Stuttgart · New York · ISSN 0032-0943

sotrine, with a 3*R*,5*S* absolute configuration, and was identified as (+)-10,11-dioxoerysotrine.

The EIMS of compound 2 showed a molecular ion peak, [M]+, at m/z 286.1, consistent with the molecular formula $C_{16}H_{14}O_5$. The ¹H- and ¹³C-NMR data (Table 1), together with COSY spectrum indicated the presence of three ring systems from the following features: 2,4,5-trioxyphenyl substitution (δ_H = 7.04, d, J = 0.8 Hz and $\delta_{\rm H}$ = 7.10, d, J = 0.8 Hz, 3,5-dioxyphenyl substitution $(\delta_{\rm H}$ = 6.95, dd, J = 2.2, 2.2 Hz; $\delta_{\rm H}$ = 6.92, dd, J = 2.2, 2.2 Hz and $\delta_{\rm H}$ = 6.41, t, J = 2.2 Hz) and a singlet at $\delta_{\rm H}$ = 7.14 ($\delta_{\rm C}$ = 102.8) characteristic of an H-3 proton of a 2-phenylbenzofuran moiety. The ¹H-NMR spectrum further showed two methoxy groups at $\delta_{\rm H}$ = 3.91 and = 3.84 which were placed at C-5 and C-3′, respectively, of a 2-phenylbenzofuran nucleus using the observed HMBC correlations between these methoxy signals and the corresponding carbon atoms. The rest of the assignments were done using HMBC data, enabling 2 to be identified as 2-(5'-hydroxy-3'methoxyphenyl)-6-hydroxy-5-methoxybenzofuran, a new natural product and also the first report of a phenylbenzofuran derivative in Erythrina species. The seed pods and the root bark were toxic to brine shrimp (Artemia salina) giving LC₅₀ values of 6.31 and 31.61, respectively, while 1 gave a value of 12.61. Compound **2** (Table **2**) exhibited the strongest antimicrobial activity against all test microorganisms while the other pure compounds 1, erysotrine, erysodine, syringaresinol and vanillic acid were inactive. The pure compounds from the root bark exhibited antimicrobial activity with the pterocarpans, isoneorautenol 3, shinpterocarpin 4, 9-0-acetylshinpterocarpin 5, erybraedin A, neorautenol and 2',4'-diacetyl-6",6"-dimethylpyrano[2",3":7,8]isoflav-3-ene **6**, showing strong activity against the yeast spores but no activity against the Gram-negative bacteria (Table 2). Previous antimicrobial activity evaluation has only been done on erybraedin A [8] and abyssinone IV [10].

Materials and Methods

General experimental procedures: M.p's uncorrected-: Optical rotations [α]_D: Polatronic-D (Schimdt + Haensch) polarimeter. UV: Shimadzu UV-2501PC Spectrophotometer. IR: Perkin Elmer 2000 FT-IR Spectrometer. 1 H-NMR, 13 C-NMR, DEPT, COSY, HMQC, HMBC: Bruker Avance DPX 300 Spectrometer using standard pulse sequences and referenced to residual solvent signal. HR-

	•		-	
	$\delta_{\!\scriptscriptstyle H}$	δ_{C}	$\delta_{\!\scriptscriptstyle H}$	δ_{C}
1	6.74, dd, (11.5,2.4)	124.9 (d)		
2	6.00, d, (11.5)	132.6 (d)		159.2 (s)
3	3.64, t, (7.9)	76.1 (d)	7.14, s	102.8 (d)
4	2.30, d, (7.9)	49.9 (t)	7.10, d, (0.8)	102.5 (d)
5		70.8 (s)		145.8 (s)
6		138.0 (s)		146.1 (s)
7	5.88, br s	121.2 (d)	7.04, d, (0.8)	97.9 (d)
8	4.64, br s	54.7 (t)		150.2 (s)
9				121.0 (s)
10		159.7 (s)		
11		181.8 (s)		
12		124.2 (s)		
13		141.7 (s)		
14	7.17, s	106.4 (d)		
15		153.5 (s)		
16		149.5 (s)		
17	7.48, s	111.1 (d)		
3-OCH₃	3.23, s	56.9 (q)		
15-OCH ₃	3.91, s	56.7 (q)		
16-OCH ₃	3.95, s	56.7 (q)		
1′				133.0 (s)
2′			6.92, dd, (2.2, 2.2)	101.5 (d)
3′				161.8 (s)
4′			6.41, t, (2.2)	101.5 (d)
5′				154.8 (s)
6′			6.95, dd, (2.2, 2.2)	104.1 (d)
5-OCH₃			3.91, s	55.3 (q)
3′-OCH₃			3.84, s	55.1 (q)

¹H- and ¹³C-NMR data for compounds **1**^a and **2**^b

1

Letter... Planta Med 2002; 68: 640 – 642

EIMS: Autospec Time of Flight (TOF) Spectrometer. EI-MS: Finnigan MAT SSQ 7000 Single Quadrupole Instrument at 70 eV. CC: Silica gel 60 (0.040-0.063 mm, Merck). Preparative and analytical TLC: silica gel 60 PF₂₅₄₊₃₆₆ (Merck). Visualisation of chromatograms: UV (254 and 366 nm) and vanillin-sulphuric acid spray.

The plant material was collected in Mapoka (Botswana) and was identified by Dr. L. M. Turton and a voucher specimen (E 0897) deposited at the University of Botswana Herbarium.

Extraction and isolation: Seed pods (5 kg) were powdered and extracted in 1:1 MeOH/CHCl₃ mixture (10 L). Removal of solvent from the extract gave a green residue (40 g) which was re-dissolved in CHCl₃/MeOH mixture (1 L) and 300 g of activated charcoal added to remove chlorophyll. The mixture was filtered and the filtrate, upon solvent evaporation, yielded a brown residue (27 g) which was chromatographed on silica gel 60 column (300 g) using a 6:1 CHCl₃/EtOAc mixture. A total of 25 fractions (200 mL each) were collected and monitored using TLC to give SPA (frs 1-17, 5 g) and SP_B (frs 18-25, 2 g). Fraction SP_A was further purified on preparative TLC (CHCl₃/EtOAc, 6:1) by multiple development (\times 3) to give (+)-10,11-dioxoerysotrine 1 (50 mg, R_f 0.32), ervsotrine (10 mg, R_f 0.58) [5], erysodine (5 mg, R_f 0.21) [5] and syringaresinol (15 mg, R_f 0.41) [11] while fraction SP_B gave 2-(5'-hydroxy-3'-methoxyphenyl)-6-hydroxy-5-methoxybenzofuran 2 (10 mg, R_f 0.19) and 4-hydroxy-3-methoxybenzoic acid (5 mg, R_f 0.30). The root bark (1.5 kg) was treated similarly to yield erybraedin A (8 mg, R_f 0.61) [8], neobavaisoflavone (120 mg, R_f 0.35) [12], [13], abyssinone IV (90 mg, R_f 0.52) [10], neorautenol (9 mg, R_f 0.40) [14], isoneorautenol (10 mg, R_f 0.48) [8], [9], shinpterocarpin (90 mg, R_f 0.35) [15] and erythrinin B (20 mg, R_f 0.29) (16). Acetylation of shinpterocarpin at room and elevated temperature gave pure **5** (13.5 mg, R_f 0.65) and **6** (22 mg, R_f 0.81), respectively. The brine shrimp lethality assay [17] was used to assess extracts and pure compounds. TLC bioautography for antibacterial and antifungal assays [18] were done with chloramphenicol and miconazole, respectively, as positive standards.

(+)-10,11-Dioxoerysotrine (1): Brown solid (acetone), m.p. 174 – 176 °C; [α] 25 : + 167.5° (MeOH, c 0.10, 25 °C); HR-TOF-MS: m/z = 341.1265 (calcd for C₁₉H₁₉NO₅, 341.1264); EI-MS: m/z (%) = 341 [M] $^+$ (100), 310 (30), 282 (25), 257 (20); IR: $v_{\rm max}^{\rm KBr}$ = 1710, 1680, 1650, 1610 cm $^-$ 1; UV: $\lambda_{\rm Max}^{\rm MeOH}$ (log ε) = 351 (3.54), 292 (sh, 3.62), 247 (4.17), 206 (4.38) nm. 1 H- and 13 C-NMR spectra data (see Table 1).

2-(5'-Hydroxy-3'-methoxyphenyl)-6-hydroxy-5-methoxybenzofuran (**2**): Brown paste (ethyl acetate), m.p., 82 – 85 °C. EI-MS: m/z (%) = 286.1 [M]+ (100), 271.0 (60), 240.9 (25); IR: $v_{\text{max}}^{\text{KBr}}$ = 3395, 2924, 1593, 1450 cm⁻¹; UV: $\lambda_{\text{Max}}^{\text{HeOH}}$ (log ε) = 324 (4.21), 293 (4.04); 285 (3.74), 216 (3.45); +NaOMe 342, 206; +NaOAc 397, 387, 324, 293, 206; +NaOAc/H₃BO₃ 397, 391, 387, 324, 293, 206; +AlCl₃ 324, 294, 215; +AlCl₃/HCl 324, 293, 255, 213 nm. ¹H- and ¹³C-NMR spectra data (see Table **1**).

Table 2 Antimicrobial activity of pure compounds from Erythrina latissima

Microorganism	Minimum inhibition amounts (μg)									
	2	3	4	5	6	7	8	9	10	11
E. coli (NCTC09001)	0.05	-	-	-	-	-	-	-	10.0	90.0
P. aeruginosa (NCTC10332)	0.05	-	-	-	-	-	-	-	0.80	90.0
S. aureus (NCTC8582)	0.008	1.00	0.70	0.70	0.80	0.55	1.25	0.50	0.40	0.50
B. subtilis (NCTC3610)	0.005	1.00	1.00	1.00	0.90	0.50	1.20	0.30	0.50	0.60
C. mycoderma (SM428)	0.001	0.008	0.008	0.01	0.01	0.007	0.01	8.00	0.06	0.07
S. cerevisiae (SM10716)	0.001	0.009	0.007	0.008	0.008	0.006	0.01	6.00	0.06	0.06

^{7:} Erybraedin A; 8: neorautenol; 9: abyssinone IV; 10: erythrinin B; 11: neobavaisoflavone.

^a In chloroform-d.

b In acetone-d₆

642

Acknowledgements

C. C. W. W. thanks (DAAD), Germany through UNESCO-ROSTA for a Scholarship and Jomo Kenyatta University of Agriculture and Technology for a study leave and R. R. T. M. thanks IFS and UBRPC for research grants. Dr. L. M. Turton (retired botanist/chemist, University of Botswana) is acknowledged for identifying the plant.

References

- ¹ Joubert FJ. Purification and properties of proteinase inhibitors from *Erythrina corallodendron* seeds. Phytochemistry 1988; 27: 1297 300
- ² Barron D, Ibrahim RK. Isoprenylated flavonoids A survey. Phytochemistry 1996; 43: 921 82
- ³ Dewick PM. The flavonoids: Advances in Research since 1986. J.B. Harborne, editor. New York: Springer, 1994: 117
- ⁴ Van-Wyk B, Van-Wyk P. Field guide to trees of Southern Africa. Cape Town: Struik Publishers, 1997: 390
- ⁵ Amer ME, Shamma M, Freyer AJ. The tetracyclic *Erythrina* alkaloid. Journal of Natural Products 1991; 54: 329–63
- ⁶ Tanaka H, Tanaka T, Etoh H, Goto S, Terada Y. Two new erythrinan alkaloids from *Erythrina* x *bidwilli*. Heterocycles 1999; 51: 2759 – 64
- ⁷ Amer M E, El-Masry S, Shamma M, Freyer A J. Three novel glycodienoid alkaloids from *Erythrina lysistemon*. Journal Natural Products 1991; 54: 161–6
- Mitscher LA, Okwute SK, Gollapudi SR, Drake S Avona E. Antimicrobial pterocarpans of Nigerian *Erythrina mildbraedii*. Phytochemistry 1988; 27: 3449 – 52
- ⁹ Kitagawa I, Chen WZ, Hori K, Harada E, Yasuda N, Yoshikawa M, Ren J. Chemical studies of Chinese Licorice-roots. I. Elucidation of five new flavonoid constituents from the roots of *Glycyrrhiza glabra* L. collected from Xinjiang. Chemical and Pharmaceutical Bulletin 1994; 42: 1056–62
- ¹⁰ Kamat VS, Chuo FY, Kubo I, Nakanishi K. Antimicrobial agents from an East African medicinal plant *Erythrina abyssinica*. Heterocycles 1981; 15: 1163-9
- ¹¹ Abe F, Yamauchi T. 9α-Hydroxypinoresinol, 9α-hydroxymedinoresinol and related lignans from *Allamanda neriifolia*. Phytochemistry 1988; 27: 575 – 7
- ¹² Nkengfack AE, Vouffo TW, Fomum ZT, Meyer M, Bergendorff O, Sterner O. Prenylated isoflavanone from the roots of *Erythrina sigmoidea*. Phytochemistry 1994; 36: 1047 51
- ¹³ Saxena VK, Bhadoria BK. 3'-Prenyl-4'-methoxy-isoflavone-7-β-D-(2"-O-ρ-coumaroyl)-glucopyranoside, a novel phytoestrogen from *Sopobia delphinifolia*. Journal of Natural Products 1990; 53: 62 5
- ¹⁴ Brink AJ, Rall GJ, Engelbrecht JP. Structures of some minor pterocarpans of *Neorautanenia edulis*. Phytochemistry 1974; 13: 1581 5
- ¹⁵ Linuma M, Ohyama M, Tanaka T. Flavonoids in roots of Saphora prostrata. Phytochemistry 1995; 38: 539-43
- ¹⁶ Deshpande VH, Pendse AD, Pendse R. Erythranins A, B and C, three new isoflavones from the bark of *Erythrina variegata*. Indian Journal of Chemistry B 1977; 15: 205 – 8
- ¹⁷ McLaughlin JL, Rogers L. The use of biological assays to evaluate botanicals. Drug Information Journal 1998; 32: 513–24
- ¹⁸ Rahalison L, Hamburger M, Hostettman K, Monod M, Frenk E. A bioautographic agar overlay method for detection of antifungal compounds from higher plants. Phytochemical Analysis 1991; 2: 199 203